**Introduction:**

Natural disturbances, including fires, windstorms, floods, droughts, and insect outbreaks, have occurred in forests for millions of years (Lindenmayer et al. 2012). In forests of the eastern United States, wind disturbance from hurricanes, derechos (straight-line windstorms), and tornados is a dominant disturbance regime (Fischer et al. 2013). When windstorms knock down canopy trees, the increased sunlight reaching the understory and forest floor can induce environmental changes. Soil temperature extremes can increase, moisture can change depending on reduced evapotranspiration but increased summer temperature, leaf litter depth can decrease due to faster decomposition, and understory plants increase their growth rates (Greenberg and Forrest 2003, Urbanovi et al. 2014, Barber and Widick 2017). The growth of understory plants interacts with tree mortality and the creation of canopy gaps to alter microclimatic conditions (Perry et al. 2018). The fallen canopy trees cause an influx of woody debris, including both coarse and fine woody debris. Uprooted trees create tip-up mounds that alter topography of the forest floor (Perry and Herms 2019). While wind disturbance changes canopy cover and abiotic conditions, it also leaves behind many living trees, understory shrubs and herbaceous plants, seeds, root systems, soils, and surviving animals. These elements left intact after a disturbance are called biological legacies, and they influence short- and long-term changes in forest structure and function (Lindenmayer et al. 2012).

Harvesting the fallen trees after a natural disturbance is a practice called salvage-logging, and this management practice is a common response to windstorms (Lindenmayer et al. 2012). Salvage logging can help landowners recover the economic value of the fallen trees, or it may be motivated towards reducing fire risk, insect outbreak risk, or safety hazard (Perry and Herms 2019). However, there is a growing interest in managing forests in ways that conserve biodiversity, including insect populations that are threatened globally due to environmental change (Wagner 2019). This includes understanding how management practices such as salvage-logging impact insect biodiversity (Thorn et al. 2018). For example, salvage-logging alters the biological legacies left by windstorms by reducing the amount and diversity of woody debris, as well as by disturbance of understory plants, potential for soil compaction from machinery, and modified tree species composition (McNabb et al. 2001, Curtze et al. 2018, Slyder et al. 2020). The combination of natural wind disturbance followed by salvage-logging disturbance could cause longer-lasting changes to the ecosystem.

To understand how insect biodiversity is affected by forest management, an indicator taxon can be used, which is a starting point for characterizing the response of the insect community (Langor and Spence 2006). Ground beetles (Coleoptera: Carabidae) are useful indicators because they are taxonomically well known, sensitive to abiotic and biotic conditions, and sensitive to forest disturbance (Koivula 2011). Ground beetles are diverse in multiple habitats, including both mature forests and early successional habitats such as tallgrass prairies, agricultural fields, urban areas, clearcuts, and floodplains (Silverman et al. 2008, Lambeets et al. 2008, Lundgren and McCravy 2011). Within mature forests, ground beetle communities can differ based on predominate tree species, managed vs. old-growth forest, and forests with dense vs open ground vegetation (Werner and Raffa 2000, Perry et al. 2018). Thus, the occurrence and species composition of ground beetles can indicate fine-scale differences in habitat.

Two processes occur to the ground beetle community after a forest disturbance such as a windstorm or logging: influx of new species and decline of some resident species. After canopy gaps open and early successional plants begin to grow, a guild of open-habitat ground beetles typically disperses to the disturbed area. This group, including certain species of *Amara*, *Anisodactylus*, *Harpalus*, and *Chlaenius*, may immediately increase following disturbance, or may take a few years to locate the site, depending on landscape structure (Sklodowski and Garbalinska 2011, Lee et al. 2017, Barber and Widick 2017). While open-habitat species increase after disturbance, there is also a decline of forest-adapted ground beetles. For example, in Minnesota, jack pine stands that were wind-disturbed or salvaged-logged 1-3 years prior had lower numbers of *Pterostichus pensylvanicus* LeConte, *Pterostichus coracinus* (Newman), and *Sphaeroderus lecontei* Dejean than undisturbed sites (Gandhi et al. 2008). In a pine forest in Poland, forest-specialists decreased in proportional abundance over a six-year period following a tornado (Sklodowski and Garbalinska 2011). These observed decreases in forest-adapted ground beetles after windstorms and salvage-logging could be caused by a variety of factors, including environmental changes such as increased sunlight, increased summer soil temperature, and decreased leaf litter (Greenberg and Forrest 2003). Although many microclimate variables stabilize after multiple years of tree regeneration following a disturbance, other variables, such as woody debris deposition or removal, remain for decades (Gore and Patterson III 1986, Perry and Herms 2019). Studies of clearcuts in Alberta indicate that differences in beetle communities between mature and clearcut forests remain even after 27 years (Pohl et al. 2007). Thus, it is unclear how long it could take for forest-adapted ground beetles to return to areas impacted by windstorms and salvage-logging, and this subject deserves further study to inform salvage-logging practices that may increase with climate change.

While much is known about how the taxonomic composition of ground beetle communities changes after forest disturbance, less is known about how these disturbances may impact the functional diversity of beetles via changes in traits. Studies have documented that ground beetle species common in tornado-disturbed forests were smaller in size, incorporated plant material or seeds into their diets, and were capable of flight, compared to those characteristic of undisturbed forest (Sklodowski and Garbalinska 2011, Perry and Herms 2019). However, ground beetle traits can vary even among, for example, equally-sized, carnivorous, flight-incapable species. For instance, ground beetles exhibit trait syndromes (Fountain-Jones et al. 2015) associated with locomotion strategy (Forsythe 1991), including surface-walking, pushing through leaf litter, burrowing, and climbing plants (Larochelle and Larivière 2003). Separately, ground beetle species also differ in their soil moisture preference, and this trait plays an important role in habitat selection (Thiele 1977). Because forest disturbance can affect leaf litter depth, ground vegetation height, soil density, woody debris cover, and soil moisture, these habitat changes could disproportionately increase the fitness of ground beetle species with certain locomotion strategies and moisture preferences. Research is needed to determine if the effect of forest disturbance on ground beetles is mediated by their species-specific traits.

Our objective is to use ground beetles to study the long-term impacts of salvage-logging after a tornado. To do this, we compared the taxonomic and functional diversity of ground beetle communities among undisturbed forest (hereafter “forest”), unsalvaged windthrow (hereafter: “windthrow”), and salvage-logged windthrow (hereafter: “salvaged”) treatments, at three and ten years after a tornado. (1) We predicted that the taxonomic alpha-diversity would be higher for windthrow and salvaged plots than forest plots after only three years, due to the immigration of open-habitat species and the persistence of some forest species. (2) After ten years, we predicted that the taxonomic alpha-diversity of the salvaged plots would be lower than that of the windthrow plots, because the lack of woody debris in salvaged plots would hinder the reestablishment of forest-adapted species. (3) Mirroring taxonomic diversity, we expected that functional alpha-diversity would increase in salvaged and windthrow plots after three years, but that by ten years, the functional diversity of salvaged plots would be lower than that of windthrow plots, because the reduced number of habitat niches in salvaged plots would reduce the diversity of locomotion strategies and soil moisture preferences. (4) We predicted that windthrow plots would be intermediate in taxonomic and functional community composition between salvaged and forest plots, due to having less-severe canopy and understory disturbance than the salvaged plots. (5) We predicted that salvaged plots after three years would support many species of ground beetles with proportionally larger eyes, proportionally shorter antennae, and higher water affinity, traits which increase fitness in the high sunlight, high soil moisture habitat that occurs after logging.

**Methods**

Study site

Research was conducted at Powdermill Nature Preserve (PNR) in Rector, Westmoreland County, Pennsylvania (latitude: 40.159806558020556, longitude: -79.27176866978374). This preserve was established as the field research station for the Carnegie Museum of Natural History in 1956 and is largely temperate deciduous forest. The annual precipitation for the years 2012-2022 was between 45 and 70 in. with a mean of 53 in. (weather station: USC00362183) (“National Centers for Environmental Information: Past Weather” n.d.). In June 2012, a tornado uprooted or snapped many canopy trees in two large areas of the forest, each about 120 × 480 m (Figure 1). These two areas are on north- or northwest-facing slopes, which were dominated by maple (*Acer spp*.), tuliptree (*Liriodendron tulipifera*), and black cherry (*Prunus serotina*) (Murphy et al. 2015), with an understory of predominately spicebush (*Lindera benzoin*) (Calinger et al. 2015). The tornado created patchy areas of canopy openness: the impacted areas had canopy openness values of 25% up to 90% (Slyder et al. 2020). The elevation of the impacted area ranges from around 1650 ft. to 1750 ft. A waterway, Laurel Run, is found along the west side of the impacted areas.

A map of a mountain

AI-generated content may be incorrect.

Figure 1. Map of the pitfall trap locations (need to add shapefiles of impacted areas as well as a legend)

From mid-summer through winter of 2013, half of each wind-disturbed area was salvage-logged using heavy machinery to remove both fallen and residual standing trees. In 2015, three transects were established across each area of forest impacted by the tornado (total 6 transects) (Figure 1). Transects were established across the windthrow and salvaged disturbances that extended 50 m into the surrounding undisturbed forest on each side. Along each transect, four plots were established: one plot in windthrow, one in salvaged, and two in the surrounding undisturbed forest. This resulted in a sample size of 24 plots, wherein all data collection occurred.

Ground-dwelling invertebrate sampling

Ground-dwelling invertebrates were sampled using barrier pitfall traps in 2015 and 2022, representing three and ten years post-tornado (two and nine years post-salvage). Barrier pitfall traps consisted of two pairs of plastic cups (each pair having an inner 500 mL cup and an outer 1 L cup) which were placed into the ground so that the lip of the cup was flush with the ground surface. The two pairs of cups were placed 1 m from each other, and garden edging (Suncast® eco edge) was placed between them to create a barrier. Cups were filled 4 cm high with propylene glycol (recreational vehicle and marine antifreeze, Peak Company Old World Industries, Clear Lake, Texas) with a few drops of detergent. Masonite board (100 cm2) was placed at 3 cm above each cup to prevent flooding from rain. Steel hardware cloth was secured over cups using 30 cm stakes to limit mammal disturbance.

Pitfall trap sampling was conducted continuously over the growing seasons in 2015 and 2022. Trap catch was collected every two weeks, and cups were refilled with propylene glycol. In 2015, pitfall traps were installed on 27-28 May, and samples were collected on 9-10 June, 24-25 June, 8 July, 22 July, 5 August, and 17 August. In 2022, traps were installed on 1-2 June, and samples were collected on 15 June, 29 June, 13 July, 27 July, 11 August, 23 August, and 9 September. Between 2015 and 2022, plot 63 had to be moved by 27 m because of fallen debris, but the new location was still within the windthrow treatment. Trap catch was collected by pouring the sample through a fine mesh strainer and storing the contents in a specimen cup with 70% ethanol until sorting and identification.

Ground beetles (Carabidae) were identified to species using taxonomic keys (Lindroth 1961, Freitag 1969, Bousquet 2010, Bousquet and Messer 2010, Hunting 2013, Harden and Guarnieri 2017). Nomenclature followed Bousquet (2012). Species vouchers were deposited at the C. A. Triplehorn Insect Collection (OSUC), Museum of Biological Diversity, The Ohio State University, Columbus, Ohio where each specimen was given a unique identifier label (Table S1).

Trait measurements

We selected eight morphological traits of beetles that have previously been shown to relate to habitat (Table \_\_\_\_) (Fountain-Jones, Baker, and Jordan 2015). These traits are body length, antenna length, eye protrusion, eye length, pronotum width, abdomen width, rear leg length, and rear trochanter length. Traits were measured under a dissecting microscope using an ocular micrometer to the nearest 0.1 mm. For each species, traits were measured on up to six individuals, three males and three females (Fountain-Jones, Baker, and Jordan 2015), although we did not achieve this number for a few rare species. The six individuals were chosen in a way that attempted to encompass the intraspecific variation in body size observed for the species. Most (185/202) specimens used for trait measurement were collected at Powdermill Nature Reserve, but a few (17/202) measured specimens were collected in Erie and Cuyahoga counties, Ohio (Table S1). These specimens were used to supplement the sample size when less than six measurable individuals were collected at Powdermill. To control for variation in beetle body size, relative measurements of all morphological traits were calculated as their ratio to body length for each individual (Ribera et al. 2001).

In addition to measured traits, we utilized the literature to provide information for three additional traits: flight capability, water affinity, and forest affinity (Larochelle and Larivière 2003). Flight capability was coded as 1 if the species is flight-capable, 0 if the species is flight-incapable, and 0.5 if the species exhibits wing dimorphism. Water affinity was coded as 0 for xerophilous species, 1 for hygrophilous species, and 0.5 for intermediate species. Water affinity was treated as a physiological trait, because ground beetle species often have consistent humidity preferences in behavioral studies (Thiele 1977). Forest affinity was coded as “forest-specialist,” “open-habitat,” or “eurytopic” (meaning the species is found in forest clearings or in both forest and open habitats). We treated forest affinity as an ecological performance trait (following the terminology of Fountain-Jones et al. (2015)), and chose to exclude it from calculations of alpha- and beta- functional diversity.

**Table \_\_\_\_\_**. Morphological traits and literature-based traits used in this study. Forest affinity (the last trait) was excluded from calculations of alpha- and beta-functional diversity.

|  |  |
| --- | --- |
| **Trait** | **Connection to habitat** |
| Body length | Shorter body length was found for ground beetles caught in wind-disturbed forests, relative to undisturbed forests (Sklodowski and Garbalinska 2011). Body length is correlated with many other morphological traits (Barton et al. 2011). |
| Antenna length | Tactile hunter species, which rely on sense of touch more than vision, tend to have longer antennae (Bauer and Kredler 1993). Longer antenna length relative to body length was found for ground beetles caught under a tree, versus in the open (Barton et al. 2011). |
| Eye protrusion | We define eye protrusion as the difference between outer eye distance and inner eye distance, where the viewer looks downwards at the dorsal side of the beetle’s head, and the outer eye distance is the distance between the outer edges of each eye, and the inner eye distance is the distance between the inner margins of each eye. A greater eye protrusion was found in a tree-climbing ground beetle, and it may allow partial overlap in the frontal visual field. However, greater eye protrusion might prevent a ground beetle from moving through thick vegetation or soil (Talarico et al. 2007). |
| Eye length | Diurnal ground beetle species and/or those adapted to open environments tend to rely on vision for predator avoidance or prey detection (Talarico et al. 2007), and thus might have longer eyes. |
| Pronotum width | A proportionally wider pronotum can be found in robust-bodied beetles, which tend to be found within open habitats (Barton et al. 2011). A narrow pronotum can be an adaptation to reaching prey within hard-to-reach crevices or shells. A narrow pronotum may also be related to the beetle having an unobstructed view behind its eyes (Forsythe 1991). |
| Abdomen width | Similar pattern to pronotum width, with species having proportionally wider abdomens tending to be found in open habitats (Barton et al. 2011). |
| Rear leg length | Open habitats seem to favor ground beetle species with shorter legs relative to body length (Barton et al. 2011). Ground beetles with longer legs may be weaker at pushing through dense substrates (Forsythe 1991). |
| Rear trochanter length | The rear trochanter connects to the femur of the rear leg. Ground beetles have a bean-shaped rear trochanter that allows them to move between narrow crevices between bark, leaf litter, or soil. The muscle in the rear trochanter allows the rear leg to create a force in the dorsal direction, which enlarges the space and allows the beetle to move through constricted areas. The trochanter is longer, on average, in species that push themselves through soil and underneath leaf litter. It is shorter in species that walk or run above the surface of the substrate (Forsythe 1991, Talarico et al. 2007). |
| Flight capability | Flight-capable species (macropterous and with fully developed flight musculature) can exploit patchy, temporary habitats. Conversely, flight incapable species may have higher fitness within stable habitats (Ribera et al. 2001, Venn 2016). |
| Water affinity | Habitats vary in the saturation of the substrate with water, and water preference varies between ground beetle species, with some species found near riverbanks or other bodies of water, others found in moist leaf litter, and others found in dry soil. A preference for low humidity may be related to overwintering within tree stumps and logs, versus in the soil (Thiele 1977). |
| Forest affinity | Some ground beetle species tend to be caught in forests, underneath trees, whereas other species tend to be caught in fields, prairies, pastures, and other open habitats (Silverman et al. 2008). |

Forest floor environment

Environmental variables on the forest floor were quantified to assess differences among windthrow, salvaged, and undisturbed forest. Percentage canopy openness was measured using a spherical crown densiometer directly above the pitfall traps to assess light availability on the forest floor. Canopy openness was measured on 9-10 June and 5 August in 2015, and on 1-2 June in 2022. Percentage cover of ground-level vegetation, leaf litter, bare ground, fine woody debris (<10 cm diameter at the large end), coarse woody debris (≥ 10 cm diameter), and rocks were estimated in two randomly selected 1 m2 quadrats around each pitfall trap. Understory vegetation height (m) was also measured in the quadrats. Ground cover estimates were collected on 9 June, 7 July, and 5 August in 2015, and on 1-2 June, 13 July, 11 August, and 6 September in 2022. Values from the two quadrats around each pitfall trap were averaged together for a site-level mean. Soil moisture was measured at three locations adjacent to each pitfall trap using a Dynamax Inc. (Houston, Texas) TH20 portable soil moisture meter with a Theta Probe ML2x sensor. Soil moisture measurements were taken biweekly when pitfall samples were collected. The three readings were averaged together for a single mean at each plot-date combination.

Statistical analysis

Data standardization:

Ground dwelling invertebrates vary in how much they move across the forest floor, and movement may be affected by forest management (Perry et al. 2021). Pitfall traps preferentially collect insects that are more active and mobile, and consequently the number of ground beetles caught in pitfalls is reported as activity-abundance, which emphasizes that insect sampling methods have inherent biases towards certain taxa (Gandhi et al. 2008).

To determine if our sampling effort was adequate to understand the ground beetle fauna at the site, we used species accumulation curves (SACs) and Chao estimators. For each year and treatment, we created an SAC with number of sampled plots in the x-axis. This was implemented using the *specaccum* function in the R package ‘vegan’ with the ‘random’ setting, which finds the mean SAC by permuting the order of the plots (Oksanen et al. 2024, R Core Team 2024). We also created SACs using the “collector” setting in order to investigate the accumulation of ground beetle species over the course of the season (from June to August or September). To estimate a lower bound on the true species richness of ground beetles, we used an asymptotic approach (Chao and Chiu 2016). We used the Chao1 estimator, which is a nonparametric estimator that gives a lower bound on the true species richness. This estimator incorporates the number of singletons and doubletons to estimate the number of undetected species and was implemented using the function “ChaoSpecies” in the R package “SpadeR” (Chao et al. 2016).

Before doing further analyses, we accounted for occasional loss of trap catch due to animal disturbance. For each species-plot combination, we divided the count by the number of days that the pitfall trap at that plot was operational (Sklodowski and Garbalinska 2011). Thus, all activity-abundance data was corrected for number of operational days.

Activity-abundance

We calculated total activity-abundance of ground beetles as the sum of the activity-abundances of all species at a plot. Furthermore, we decomposed total activity-abundance into two groups: the contribution from forest specialist species, and the contribution from open-habitat and eurytopic species (Larochelle and Larivière 2003).

Taxonomic alpha-diversity

To investigate alpha-diversity at the plot level, we calculated species richness and Shannon diversity using the package “HillR” (Li 2018). Shannon diversity was calculated using the formula exp(-Σpiln(pi)), which is the same as the Hill number of order 1. This metric takes values between 1 and the species richness, depending on the degree to which the relative abundances are equal.

Functional alpha diversity

We used trait data in combination with ground beetle counts to calculate functional alpha-diversity. Before calculating functional alpha-diversity, we conducted a dimensionality reduction on the traits to eliminate redundant information held within the trait data. The eight continuous traits had already been standardized by dividing by body length. After investigating the Pearson correlation coefficients between pairs of traits using the package “corrplot” (Wei and Simko 2024), we determined that eye protrusion and eye length were highly correlated (r = 0.87) and that antenna length and rear leg length were highly correlated (r = 0.81), even after standardization of each variable to body length. To address this, we replaced *standardized eye protrusion* with *eye protrusion*:*eye length ratio*, and we replaced *standardized antenna length* with *antenna length*:*rear leg length ratio*. These changes resulted in a set of traits with correlation coefficients ≤ 0.51 between pairs. Trait measurements were averaged across individuals of a species to calculate species-specific means. To address any remaining collinearity within the numerical traits, we performed a principal components analysis (PCA) (Swenson 2014). We centered each continuous trait to a mean of 0, scaled to a variance of 1, and ran the PCA using the function “prcomp” in the R package “stats” (R Core Team 2024). We removed the species *Notiophilus aeneus* (Herbst, 1806) from the PCA analysis because inclusion of this rare species (3 individuals total collected) noticeably changed the PC axes (Table S\_\_\_\_). After running the PCA, we added *Notiophilus* back into the analysis by centering and scaling its data using the previously-calculated scaling coefficients and then projecting the values onto the PC axes using the loading values. We used the first four PC axes, which together explained 83% of the variance in the data, along with the categorical variables *Water affinity* and *Flight capability*, to calculate a Gower dissimilarity matrix between all ground beetle species in trait space using the package “FD” (Laliberte et al. 2014). The categorical variables were treated as ordinal data and the “metric” method was used to calculate dissimilarity, so that a wing dimorphic species would be counted as intermediate in its flight capability. We calculated functional alpha diversity for each plot by computing the weighted mean dissimilarity in trait space between pairwise combinations of species found at the plot. Each calculation of dissimilarity between a pair of species was weighted by the product of the relative abundances of the two species at the plot, so that more abundant species had a larger influence on functional alpha diversity (Swenson 2014). The calculation was carried out using the function *mpd* in the R package “picante” (Kembel et al. 2010).

Community-weighted mean traits

To investigate the average body proportions of ground beetles at each plot, we calculated community-weighted mean (CWM) trait values. These were calculated for the first three PC axes, as well as for *Water affinity*, *Flight capability*, and *Standardized antenna length*,using the function “functcomp” in the R package “FD” (Laliberte et al. 2014). The first three PC axes were considered, because together they explained 72% of the variation in the eight numerical traits, and because they had biological meaning for locomotion and sensory strategies.

Environmental variables

Before analyzing

To investigate the correlations between different ground-level environmental variables, we examined a matrix of Pearson correlations.

ran a principal components analysis (PCA) using the variables *canopy openness*, *soil moisture*, *vegetation height*, and *percent cover of vegetation*, *leaf litter*, *fine woody debris*, *coarse woody debris*, and *rock*. We ran separate PCAs for each year.

Models

We tested the relationship between forest management treatment (forest control, windthrow, salvaged) and plot-level metrics of ground beetle biodiversity and environmental variables using linear mixed-effects models with the R package “lme4” (Bates et al. 2015) and “lmerTest” (Kuznetsova et al. 2017). Our response variables were total activity-abundance, activity-abundance of open-habitat and eurytopic species, activity-abundance of forest-specialist species, species richness, Shannon diversity, functional alpha diversity, community-weighted mean (CWM) traits, and PCA axes of environmental variables. We created separate models for each year of sampling, 2015 and 2022. We included transect as a random effect to account for spatial structure in the data. For some of the models of activity-abundance, the variance of the residuals differed based on the fitted value (heteroscedasticity), so we transformed these response variables using f(x)=ln(x), before rerunning the model. In one case, the function g(x)=ln(x+0.1) was used due to the presence of zeros in the response variable. In some models, a singular fit was obtained, so we ran the model excluding transect as a random effect. In the species richness model, because species richness is a count variable, we used Poisson generalized linear mixed effects models with the packages “lme4” and “car” (Fox and Weisberg 2019). Pairwise comparisons between treatment groups were made using the package “emmeans” (Lenth 2024).

To further investigate differences between CWMs of the PCA axes, we ran models with CWMs of single morphological traits as response variables. Whenever we found a treatment difference for a PCA axis, we then located the primary traits that contributed to this axis, which we defined by having loading values with magnitude above 0.4. Then we calculated the CWMs for these traits and ran separate models for each.

Taxonomic beta diversity

To investigate whether windthrow plots had ground beetle communities that were intermediate between salvaged and forest plots, we used taxonomic beta diversity. Because we were more interested in differences in relative abundances of species rather than absolute abundances, and because the total activity-abundance varied substantially between plots, we performed a data standardization. We row-standardized using the “total” option in the function “decostand” within the R package “vegan” (Oksanen et al. 2024) so that the sum of the activity-abundances of each species at a plot summed to 1. Next, we calculated a dissimilarity matrix in species-space between all pairs of plots using Bray-Curtis dissimilarity with the function “vegdist”. Then, using a permutational multivariate analysis of variance (PERMANOVA) with the function “adonis2”, we tested the null hypothesis that the centroids of each treatment group were identical and that their dispersions were identical. Furthermore, we used an analysis of multivariate homogeneity of group dispersions with the “betadisper” function to test if the treatments differed in their dispersions. Beta-diversity was visualized using nonmetric multidimensional scaling, with two dimensions.

Functional beta diversity

Mirroring taxonomic beta-diversity, we examined functional beta-diversity to explore whether plots in different forest management treatments had species with different traits. We calculated distances between all pairs of plots using mean pairwise distance with the function “comdist” in the R package “picante” (Kembel et al. 2010). Mean pairwise distances were weighted by the species abundances. The PERMANOVA and analysis of multivariate homogeneity of group dispersions were performed as in taxonomic beta-diversity. Significant effects of forest management treatment were followed by pairwise comparisons using the R package “pairwiseAdonis” (Martinez Arbizu 2017).

**Results**

Species accumulation curves

In total, 55 species of ground beetle were collected, including 37 species in 2015 and 46 species in 2022 (Table \_\_\_\_). There were 28 species found in both years. Species accumulation curves for each treatment, using the “random” method, showed that forest plots had a better sampling effort than windthrow or salvaged plots, primarily because there were twice as many forest plots (Figure \_\_\_\_). Species accumulation curves, using the “collector” method, showed that in 2022, in the last two sampling intervals, between 11 August and 9 September, an additional 9 species were accumulated. In contrast, in 2015 the last two sampling intervals, between 22 July and 17 August, only accumulated one additional species (Figure \_\_\_). The Chao1 estimator for the true number of ground beetle species in 2015 was 47.1 species (95% conf. int. 39.3 - 82.2 species) while in 2022 the Chao1 estimator was 62.0 species (95% conf. int. 50.9 - 98.1 species).

Table \_\_\_\_. Species of ground beetle (Coleoptera: Carabidae) captured in 24 pitfall traps at Powdermill Nature Reserve. Sampling occurred from 27 May to 17 August, 2015, and from 1 June to 9 September, 2022. The species are numbered to match the numbers on the principal component analysis biplot in Figure \_\_\_.

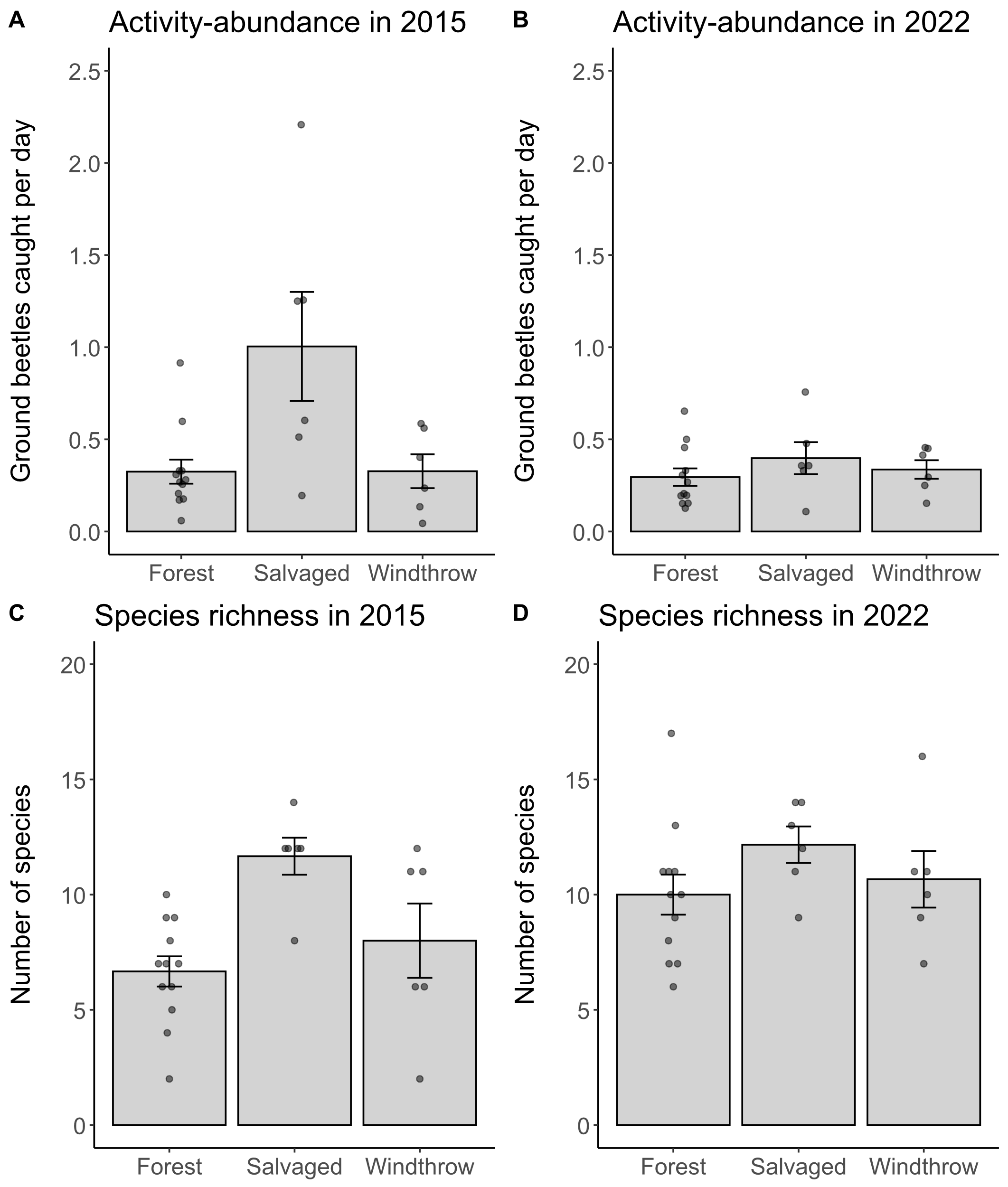
|  |  |  |  |
| --- | --- | --- | --- |
|  | Species | 2015 | 2022 |
| 1 | *Agonoleptus thoracicus* | 0 | 1 |
| 2 | *Agonum ferreum* | 1 | 0 |
| 3 | *Agonum fidele* | 31 | 3 |
| 4 | *Agonum retractum* | 1 | 0 |
| 5 | *Amerizus sp.* | 0 | 1 |
| 6 | *Amphasia interstitialis* | 6 | 0 |
| 7 | *Anisodactylus harrisii* | 2 | 1 |
| 8 | *Anisodactylus melanopus* | 1 | 1 |
| 9 | *Anisodactylus nigerrimus* | 3 | 5 |
| 10 | *Apenes lucidula* | 0 | 8 |
| 11 | *Carabus goryi* | 66 | 42 |
| 12 | *Chlaenius emarginatus* | 137 | 25 |
| 13 | *Chlaenius laticollis* | 2 | 0 |
| 14 | *Cyclotrachelus convivus* | 1 | 0 |
| 15 | *Cyclotrachelus fucatus* | 5 | 1 |
| 16 | *Cyclotrachelus sigillatus* | 22 | 53 |
| 17 | *Cymindis limbata* | 0 | 1 |
| 18 | *Cymindis platicollis* | 0 | 1 |
| 19 | *Dicaelus politus* | 54 | 17 |
| 20 | *Dicaelus teter* | 68 | 11 |
| 21 | *Galerita bicolor* | 0 | 3 |
| 22 | *Harpalus spadiceus* | 1 | 13 |
| 23 | *Lophoglossus scrutator* | 0 | 1 |
| 24 | *Myas coracinus* | 0 | 2 |
| 25 | *Notiobia nitidipennis* | 6 | 0 |
| 26 | *Olisthopus parmatus* | 3 | 2 |
| 27 | *Patrobus longicornis* | 0 | 1 |
| 28 | *Platynus angustatus* | 19 | 77 |
| 29 | *Platynus decentis* | 0 | 1 |
| 30 | *Platynus hypolithos* | 0 | 1 |
| 31 | *Platynus tenuicollis* | 3 | 6 |
| 32 | *Pseudamara arenaria* | 2 | 1 |
| 33 | *Pterostichus adoxus* | 19 | 120 |
| 34 | *Pterostichus atratus* | 0 | 2 |
| 35 | *Pterostichus coracinus* | 28 | 16 |
| 36 | *Pterostichus corvinus* | 6 | 0 |
| 37 | *Pterostichus diligendus* | 1 | 1 |
| 38 | *Pterostichus hamiltoni* | 0 | 1 |
| 39 | *Pterostichus lachrymosus* | 27 | 66 |
| 40 | *Pterostichus melanarius* | 1 | 0 |
| 41 | *Pterostichus moestus* | 185 | 47 |
| 42 | *Pterostichus mutus* | 1 | 0 |
| 43 | *Pterostichus rostratus* | 36 | 24 |
| 44 | *Pterostichus sayanus* | 0 | 2 |
| 45 | *Pterostichus stygicus* | 62 | 90 |
| 46 | *Pterostichus tristis* | 10 | 74 |
| 47 | *Scaphinotus andrewsii* | 0 | 2 |
| 48 | *Scaphinotus imperfectus* | 0 | 6 |
| 49 | *Scaphinotus ridingsii* | 0 | 2 |
| 50 | *Scaphinotus viduus* | 1 | 2 |
| 51 | *Sphaeroderus canadensis* | 35 | 7 |
| 52 | *Sphaeroderus stenostomus* | 76 | 112 |
| 53 | *Synuchus impunctatus* | 0 | 1 |
| 54 | *Trichotichnus autumnalis* | 10 | 2 |
| 55 | *Notiophilus aeneus* | 2 | 1 |
|  | Total | 932 | 856 |

Taxonomic alpha-diversity

An average of 8.3 species were found at each plot in 2015, while an average of 10.7 species were found at each plot in 2022. In 2015, species richness differed by forest management treatment (Χ2=11.967, p=0.003), with salvaged plots having higher species richness than forest plots (Z=3.426, p=0.002) (Figure \_\_\_\_). There was no significant effect of treatment on species richness in 2022 (Χ2=1.750, p=0.417). Shannon diversity also did not differ by treatment in either 2015 (F=1.064, p=0.368) or 2022 (F=0.024, p=0.976).

Total activity-abundance of ground beetles

In total, 934 individual ground beetles were caught in 2015 over 82 days of trapping, while 857 ground beetles were caught in 2022 over 112 days of trapping. On average, traps in 2015 caught 0.495 ground beetles per day, while traps in 2022 caught 0.331 ground beetles per day. Forest management treatment had a significant effect on ground beetle activity-abundance in 2015 (F=8.976, p=0.002), but not in 2022 (F=1.095, p=0.359). In 2015, salvaged plots caught more individual ground beetles than either windthrow plots (t=3.676, p=0.006) or forest plots (t=3.842, p=0.004), while forest and windthrow did not differ significantly (t=0.403, p=0.915) (Figure \_\_\_\_).



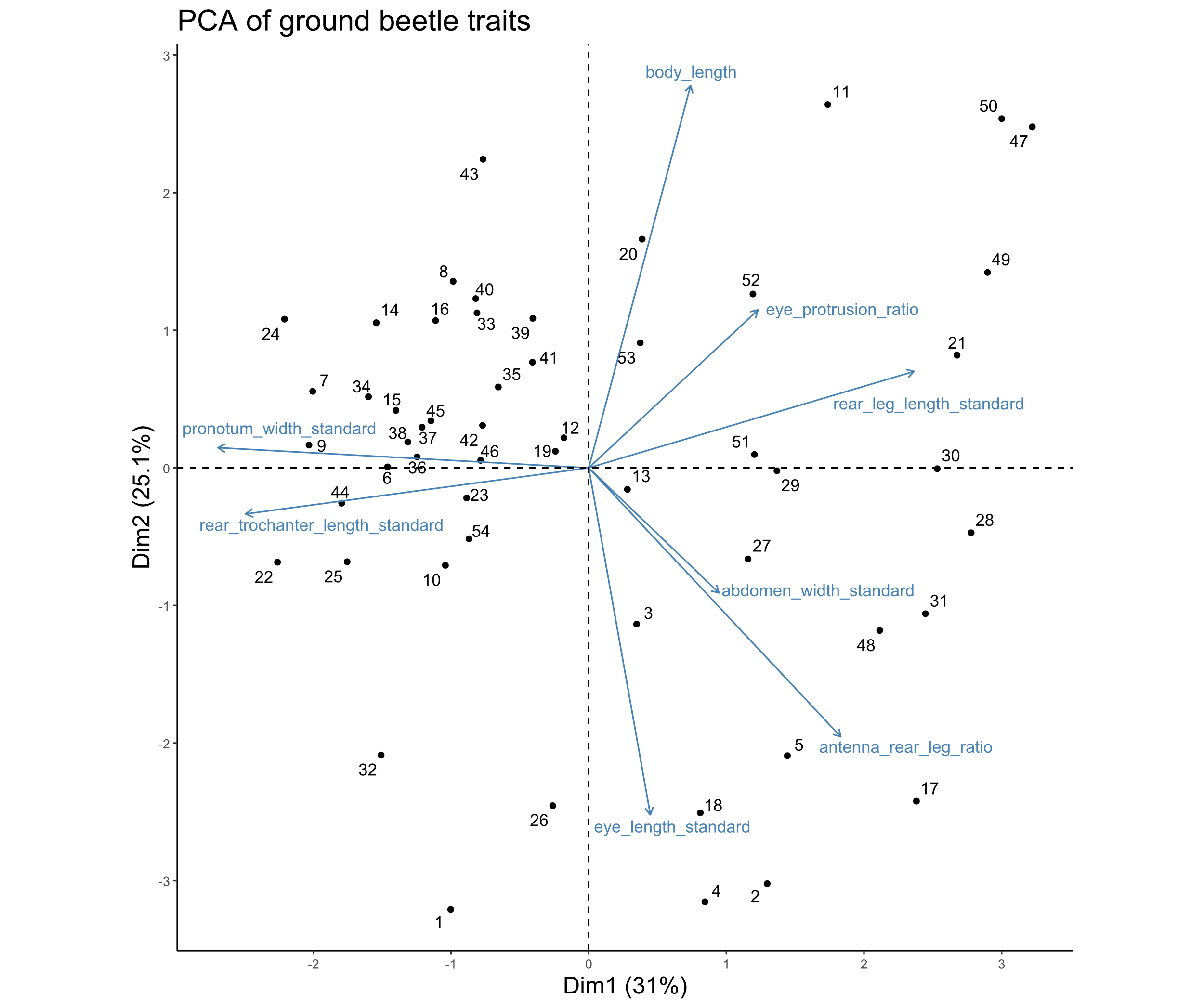
**Figure \_\_\_.** Activity-abundance and taxonomic species richness of ground beetles in 24 plots. Titles above plots indicate the year of data. Bar height is the group mean, and whiskers indicate the standard error of the mean. Points indicate the values of individual plots (ie. the ground beetles caught over one summer season at a particular location). A, B) Activity-abundance is measured in mean ground beetles caught per day. C, D) Species richness is measured in number of distinct ground beetle species captured. The 2015 trapping interval was 82 days long, while the 2022 trapping interval was 112 days long.

Activity-abundance by forest affinity

Of the 55 ground beetle species found in the study, 23 species were categorized as forest specialists, 28 species were eurytopic, 2 species were listed as open-habitat, and 2 had unclear or unknown forest affinity (Larochelle and Larivière 2003). In 2015, there was a higher activity-abundance of open-habitat or eurytopic ground beetles in the salvaged plots than in the forest plots (t=3.612, p=0.006) or windthrow plots (t=3.044, p=0.020), while in 2022 there was no significant effect (2015: F=7.211, p=0.006; 2022: F=0.836, p=0.452). There were not any significant treatment differences in the activity-abundance of forest-specialist ground beetles in either 2015 (F=2.177, p=0.138) or 2022 (F=0.127, p=0.881).

Principal components of traits

The PCA analysis of the eight numerical traits generated a set of four axes which together explained 83% of the variance. The first PC axis (31% of the variance) was associated with proportionally narrower pronotum (relative to body length), proportionally longer rear legs, and proportionally shorter rear trochanter (Figure \_\_\_\_, Table \_\_\_\_\_). The second PC axis (25% of the variance) was associated with longer body length, proportionally shorter eye length, and smaller (antenna length):(rear leg length) ratio. The third PC axis (16% of the variance) was associated with proportionally wider abdomen and larger eye protrusion ratio. The fourth PC axis (11% of the variance) was associated with proportionally longer eyes, proportionally longer rear legs, and proportionally longer rear trochanters.



**Figure \_\_\_\_.** Biplot of the first two principal component axes of the PCA of the eight numerical traits measured for 54 ground beetle species. The species *Notiophilus aeneus* was omitted from the analysis. Numbers from 1 to 54 indicate species, which are listed in Table \_\_\_\_\_. Blue arrows with labels are the eight trait variables. The word “standard” after a variable name indicates that it has previously been divided by body length to address high correlations with body length.

**Table \_\_\_\_\_.** Loading values for the principal components analysis of eight numerical traits. The word “standard” after a variable name indicates that it has previously been divided by body length to address high correlations with body length. Eye protrusion ratio is equal to eye protrusion divided by eye length. Antenna rear leg ratio is equal to antenna length divided by rear leg length.

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| Trait | PC1 | PC2 | PC3 | PC4 | PC5 | PC6 | PC7 | PC8 |
| body\_length | 0.15 | 0.61 | -0.10 | 0.27 | 0.06 | 0.19 | 0.57 | -0.39 |
| eye\_length\_standard | 0.09 | -0.55 | -0.07 | 0.52 | 0.21 | -0.37 | 0.10 | -0.46 |
| eye\_protrusion\_ratio | 0.24 | 0.25 | 0.58 | -0.02 | 0.68 | -0.17 | -0.22 | 0.03 |
| pronotum\_width\_standard | -0.53 | 0.03 | 0.36 | 0.03 | -0.12 | 0.36 | -0.35 | -0.57 |
| abdomen\_width\_standard | 0.19 | -0.20 | 0.70 | 0.25 | -0.45 | 0.11 | 0.33 | 0.23 |
| rear\_leg\_length\_standard | 0.47 | 0.15 | -0.17 | 0.51 | -0.18 | 0.30 | -0.58 | 0.07 |
| antenna\_rear\_leg\_ratio | 0.36 | -0.43 | -0.05 | -0.31 | 0.29 | 0.68 | 0.15 | -0.14 |
| rear\_trochanter\_length\_standard | -0.49 | -0.07 | -0.08 | 0.48 | 0.39 | 0.34 | 0.14 | 0.48 |

Functional alpha-diversity

The functional alpha-diversity, as measured by mean pairwise distance in trait space of species found at a plot, did not differ significantly by forest management treatment, for either 2015 (F=1.332, p=0.292) or 2022 (F=0.263, p=0.771).

Community-weighted means

The community-weighted mean (CWM) values at each plot, of the first three PC axes, were analyzed for any differences between forest management treatment groups. The CWM of PC1 varied by treatment group in both 2015 (F=4.099, p=0.031) and 2022 (F=5.119, p=0.015). Plots that were salvage-logged tended to capture ground beetles with smaller values of PC1 than undisturbed forest plots in both 2015 (t=2.763, p=0.030) and 2022 (t=3.123, p=0.014). Further investigation of the traits contributing to PC1 found treatment differences for *rear trochanter length* in 2015 (F=8.735, p=0.003) and *pronotum width* in 2022 (F=3.895, p=0.036), but no significant differences in *rear leg length*. Specifically, in 2015 ground beetles caught in salvaged plots tended to have proportionally longer trochanters than those caught in forest plots (t=4.176, p=0.002). In 2022, ground beetles caught in salvaged plots tended to have proportionally wider pronota than those caught in forest plots (t=2.584, p=0.044). These variables had been previously standardized by dividing by body length, so differences reflect body proportions rather than absolute sizes.

The CWM of PC2 also varied by treatment group in 2015 (F=8.063, p=0.004), but did not vary significantly by treatment group in 2022 (F=2.839, p=0.088). In 2015, ground beetles caught in the salvaged treatment had, on average, smaller values of PC2 than those caught in the forest treatment (t=3.993, p=0.003). Specifically, there were differences in *eye length* by treatment in 2015 (F=8.344, p=0.003), but no significant differences in *body length* (F=3.180, p=0.062) or *antenna length*:*rear leg length ratio* (F=0.022, p=0.978) for 2015. In 2015, ground beetles caught in salvage-logged locations tended to have proportionally longer eyes relative to their body length than ground beetles caught in un-harvested windthrow locations (t=2.657, p=0.043) or undisturbed forest (t=4.068, p=0.002). The CWM of PC3 did not vary significantly by treatment group for either 2015 (F=3.6096, p=0.051) or 2022 (F=0.424, p=0.661).

The CWM *Water affinity* did not vary significantly by treatment group in either 2015 (F=1.657, p=0.215) or 2022 (F=0.345, p=0.712). The CWM *Flight capability* varied by treatment in 2015 (F=7.143, p=0.006) but not in 2022 (F=1.444, p=0.265). Salvaged plots in 2015 tended to catch a higher proportion of flight-capable ground beetles than windthrow plots (F=3.085, p=0.018) or undisturbed forest plots (F=3.564, p=0.007). The CWM of *standardized antenna length* did not significantly vary by treatment group in either 2015 (F=0.450, p=0.645) or 2022 (F=2.622, p=0.096).

Table \_\_\_\_. Summary of the models used to test for differences between ground beetles in salvaged, windthrow, and forest plots. The letter superscripts after values in the Forest, Salvaged, and Windthrow columns indicate results of Tukey pairwise significance tests, with different letters indicating that the two groups differ significantly between the two forest treatments at α=0.5. The model type column indicates if the model was a standard linear model (LM), a linear mixed-effects model with Transect as a random effect (LMM), or a generalized linear mixed effects model with Poisson errors (GLMM).

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| Variable | Unit | Year | Forest | Salvaged | Windthrow | Model  type | Test statistic | p-value |
| Total activity-abundance | Beetles/day | 2015 | **0.325 a** | **1.004 b** | **0.327 a** | **LMM\*** | **F=8.976** | **0.002** |
|  |  | 2022 | 0.295 | 0.398 | 0.336 | LMM | F=1.095 | 0.359 |
| Activity abundance of open-habitat and eurytopic species | Beetles/day | 2015 | **0.155 a** | **0.551 b** | **0.152 a** | **LMM\*** | **F=7.212** | **0.006** |
|  |  | 2022 | 0.130 | 0.246 | 0.159 | LMM\* | F=0.836 | 0.452 |
| Activity abundance of forest-specialist species | Beetles/day | 2015 | 0.170 | 0.453 | 0.175 | LM\* | F=2.177 | 0.138 |
|  |  | 2022 | 0.165 | 0.151 | 0.178 | LMM | F=0.127 | 0.881 |
| Species richness | Number of species | 2015 | **6.667 a** | **11.667 b** | **8.000 ab** | **GLMM** | **X2=11.97** | **0.003** |
|  |  | 2022 | 10.000 | 12.167 | 10.667 | GLMM | X2=1.750 | 0.417 |
| Shannon diversity | Effective number of species | 2015 | 5.119 | 6.315 | 6.450 | LMM | F=1.064 | 0.368 |
|  |  | 2022 | 7.297 | 7.468 | 7.466 | LMM | F=0.024 | 0.9763 |
| Functional alpha-diversity | Within-plot mean pairwise distance in trait space | 2015 | 0.154 | 0.181 | 0.156 |  |  |  |
|  |  | 2022 | 0.173 | 0.164 | 0.171 |  |  |  |
| Community-weighted mean (CWM) principal component (PC) 1 |  | 2015 | 0.287 | -0.210 | -0.006 |  |  |  |
|  |  | 2022 | 0.297 | -0.207 | 0.023 |  |  |  |
| CWM Standardized pronotum width |  | 2015 | 0.262 | 0.265 | 0.263 |  |  |  |
|  |  | 2022 | 0.259 | 0.265 | 0.264 |  |  |  |
| CWM Standardized rear leg length |  | 2015 | 0.744 | 0.752 | 0.731 |  |  |  |
|  |  | 2022 | 0.761 | 0.730 | 0.743 |  |  |  |
| CWM Standardized rear trochanter length |  | 2015 | 0.098 | 0.113 | 0.103 |  |  |  |
|  |  | 2022 | 0.100 | 0.106 | 0.102 |  |  |  |
| CWM PC2 |  | 2015 | 1.212 | 0.486 | 0.896 |  |  |  |
|  |  | 2022 | 0.853 | 0.592 | 0.782 |  |  |  |
| CWM Standardized eye length |  | 2015 | 0.045 | 0.050 | 0.046 |  |  |  |
| CWM Standardized body length |  | 2015 | 17.323 | 15.607 | 16.093 |  |  |  |
| CWM (Antenna length):(Rear leg length) ratio |  | 2015 | 0.607 | 0.608 | 0.606 |  |  |  |
| CWM PC3 |  | 2015 | -0.272 | -0.616 | -0.446 |  |  |  |
|  |  | 2022 | -0.449 | -0.498 | -0.367 |  |  |  |
| CWM Water affinity |  | 2015 | 0.791 | 0.796 | 0.707 |  |  |  |
|  |  | 2022 | 0.758 | 0.782 | 0.797 |  |  |  |
| CWM Flight capability |  | 2015 | 0.077 | 0.297 | 0.077 |  |  |  |
|  |  | 2022 | 0.034 | 0.065 | 0.075 |  |  |  |
| CWM Standardized antenna length |  | 2015 | 0.453 | 0.458 | 0.444 |  |  |  |
|  |  | 2022 | 0.464 | 0.438 | 0.447 |  |  |  |

\*: a log transformation was done on the response variable to improve the model’s satisfaction of assumptions

Taxonomic beta-diversity

The PERMANOVA tests indicated no significant differences between the taxonomic community composition of forest, windthrow, and salvage plots for either 2015 (F=1.482, p=0.077) or 2022 (F=1.076, p=0.355). Furthermore, the analysis of homogeneity of group dispersions found that forest, windthrow, and salvage plots did not differ in dispersion in either the 2015 analysis (F=0.158, p=0.855) or the 2022 analysis (F=2.029, p=0.157). The NMDS fits for the 2015 and 2022 data had stress values of 0.150 and 0.163, respectively.

Functional beta-diversity

The PERMANOVA tests indicated that there were differences in the functional community composition of ground beetles between different forest treatments in 2015 (F=1.841, p=0.004), but no significant differences in 2022 (F=1.043, p=0.290). In 2015, salvaged plots differed from windthrow plots (F=1.837, p=0.013) and from forest plots (F=2.485, p=0.002), which indicates a difference in either centroids, group dispersions, or both. The NMDS fit for 2015 had a stress value of 0.141, while the 2022 NMDS had a stress value of 0.211.

Environmental variables

**Discussion**

Although the percent cover of woody debris did not differ between the windthrow and salvaged treatments, there was higher volume of coarse woody debris in the windthrow in 2014, compared to the salvaged treatment (Perry 2016). The reduction in coarse woody debris volume in the salvaged treatment could last decades. When examining clearcut stands of varying age in New Hampshire, researchers found that the slash from clearcutting decomposed within 20-30 years, leaving low mass of downed wood for an additional 30 years, before the tree regeneration began to contribute downed wood (Gore and Patterson III 1986). Thus, salvage-logging could affect woody debris volume for >50 years.

It must be mentioned that just because a ground beetle was caught in a given forest disturbance treatment doesn’t mean that it spent the entire year at that location. Ground beetles, even flightless species, can move fairly long distances over the seasons (cite). For example, some ground beetle species forage for prey in the summer within agricultural fields but move to woodlot edges to overwinter (cite). Thus, the fact that many forest ground beetles like \_\_\_\_ and \_\_\_\_ were still caught in salvage-logged areas does not rule out that they could still need the mature forest for part of the year.

I think it’s worth mentioning that the 2022 survey caught more species despite the total number of ground beetles caught being lower. It is likely due to the extended trapping season, which went into September.

Probably one of the more interesting results of the NMDS analysis is that the southwest area had plots with higher dispersion in community composition than the northeast area. Maybe this is due to the presence of the creek in the southwest area.

**Supplementary information**

**Table S1.** Voucher specimens used for trait measurements in this study. The Unique ID begins with “OSUC” if the specimen was already in the Ohio State University Triplehorn Insect Collection from 2015 or a previous study, whereas Unique IDs with just a number indicate 2022 specimens which will be vouchered as part of this study.

|  |  |  |  |
| --- | --- | --- | --- |
| Species | Sex | Unique ID | Location of collection |
| *Agonoleptus thoracicus* | f | 819611 | Powdermill |
| *Agonum ferreum* | m | OSUC 671705 | Powdermill |
| *Agonum ferreum* | m | OSUC 671700 | Powdermill |
| *Agonum ferreum* | f | OSUC 671704 | Powdermill |
| *Agonum ferreum* | f | OSUC 671703 | Powdermill |
| *Agonum ferreum* | f | OSUC 671702 | Powdermill |
| *Agonum fidele* | f | 819619 | Powdermill |
| *Agonum fidele* | f | 819616 | Powdermill |
| *Agonum fidele* | f | 819613 | Powdermill |
| *Agonum retractum* | m | OSUC 671697 | Powdermill |
| *Agonum retractum* | f | OSUC 523212 | Erie Co. OH |
| *Agonum retractum* | f | OSUC 523213 | Erie Co. OH |
| *Agonum retractum* | f | OSUC 523214 | Erie Co. OH |
| *Amerizus sp.* | m | 819614 | Powdermill |
| *Amphasia interstitialis* | m | Not vouchered | Powdermill |
| *Amphasia interstitialis* | f | Not vouchered | Powdermill |
| *Amphasia interstitialis* | f | Not vouchered | Powdermill |
| *Amphasia interstitialis* | f | Not vouchered | Powdermill |
| *Anisodactylus harrisii* | m | OSUC 671671 | Powdermill |
| *Anisodactylus harrisii* | m | OSUC 740908 | Cuyahoga Co. OH |
| *Anisodactylus harrisii* | m | OSUC 740906 | Cuyahoga Co. OH |
| *Anisodactylus harrisii* | f | 819778 | Powdermill |
| *Anisodactylus harrisii* | f | OSUC 740902 | Cuyahoga Co. OH |
| *Anisodactylus harrisii* | f | OSUC 740900 | Cuyahoga Co. OH |
| *Anisodactylus melanopus* | m | 819699 | Powdermill |
| *Anisodactylus nigerrimus* | m | 819634 | Powdermill |
| *Anisodactylus nigerrimus* | m | 819631 | Powdermill |
| *Anisodactylus nigerrimus* | m | 819628 | Powdermill |
| *Anisodactylus nigerrimus* | f | 819625 | Powdermill |
| *Anisodactylus nigerrimus* | f | 819622 | Powdermill |
| *Apenes lucidula* | m | 819706 | Powdermill |
| *Apenes lucidula* | m | 819715 | Powdermill |
| *Apenes lucidula* | f | 819707 | Powdermill |
| *Apenes lucidula* | f | 819716 | Powdermill |
| *Apenes lucidula* | f | 819724 | Powdermill |
| *Carabus goryi* | m | 819636 | Powdermill |
| *Carabus goryi* | m | 819639 | Powdermill |
| *Carabus goryi* | m | 819642 | Powdermill |
| *Carabus goryi* | f | 819645 | Powdermill |
| *Carabus goryi* | f | 819648 | Powdermill |
| *Carabus goryi* | f | 819651 | Powdermill |
| *Chlaenius emarginatus* | m | 819704 | Powdermill |
| *Chlaenius emarginatus* | m | 819713 | Powdermill |
| *Chlaenius emarginatus* | m | 819722 | Powdermill |
| *Chlaenius emarginatus* | f | 819705 | Powdermill |
| *Chlaenius emarginatus* | f | 819714 | Powdermill |
| *Chlaenius emarginatus* | f | 819723 | Powdermill |
| *Chlaenius laticollis* | f | OSUC 671794 | Powdermill |
| *Chlaenius laticollis* | f | OSUC 671795 | Powdermill |
| *Cyclotrachelus convivus* | m | OSUC 671699 | Powdermill |
| *Cyclotrachelus convivus* | m | OSUC 522517 | Erie Co. OH |
| *Cyclotrachelus convivus* | m | OSUC 740962 | Cuyahoga Co. OH |
| *Cyclotrachelus convivus* | f | OSUC 522528 | Erie Co. OH |
| *Cyclotrachelus convivus* | f | OSUC 522527 | Erie Co. OH |
| *Cyclotrachelus convivus* | f | OSUC 522525 | Erie Co. OH |
| *Cyclotrachelus fucatus* | m | 819782 | Powdermill |
| *Cyclotrachelus sigillatus* | m | 819753 | Powdermill |
| *Cyclotrachelus sigillatus* | m | 819763 | Powdermill |
| *Cyclotrachelus sigillatus* | m | 819773 | Powdermill |
| *Cyclotrachelus sigillatus* | f | 819754 | Powdermill |
| *Cyclotrachelus sigillatus* | f | 819764 | Powdermill |
| *Cyclotrachelus sigillatus* | f | 819774 | Powdermill |
| *Cymindis limbata* | unknown sex | 819617 | Powdermill |
| *Cymindis platicollis* | unknown sex | 819777 | Powdermill |
| *Dicaelus politus* | m | 819708 | Powdermill |
| *Dicaelus politus* | m | 819717 | Powdermill |
| *Dicaelus politus* | m | 819725 | Powdermill |
| *Dicaelus politus* | f | 819709 | Powdermill |
| *Dicaelus politus* | f | 819718 | Powdermill |
| *Dicaelus politus* | f | 819726 | Powdermill |
| *Dicaelus teter* | m | 819604 | Powdermill |
| *Dicaelus teter* | m | 819719 | Powdermill |
| *Dicaelus teter* | m | 819727 | Powdermill |
| *Dicaelus teter* | f | 819710 | Powdermill |
| *Dicaelus teter* | f | 819720 | Powdermill |
| *Dicaelus teter* | f | 819728 | Powdermill |
| *Galerita bicolor* | f | 819640 | Powdermill |
| *Galerita bicolor* | f | 819637 | Powdermill |
| *Harpalus spadiceus* | m | 819658 | Powdermill |
| *Harpalus spadiceus* | m | 819655 | Powdermill |
| *Harpalus spadiceus* | m | 819652 | Powdermill |
| *Harpalus spadiceus* | f | 819649 | Powdermill |
| *Harpalus spadiceus* | f | 819646 | Powdermill |
| *Harpalus spadiceus* | f | 819643 | Powdermill |
| *Lophoglossus scrutator* | m | 819780 | Powdermill |
| *Myas coracinus* | m | 819733 | Powdermill |
| *Myas coracinus* | f | 819734 | Powdermill |
| *Notiobia nitidipennis* | m | 672665 | Powdermill |
| *Notiobia nitidipennis* | m | 672659 | Powdermill |
| *Notiobia nitidipennis* | m | 672651 | Powdermill |
| *Notiobia nitidipennis* | f | 672663 | Powdermill |
| *Notiobia nitidipennis* | f | 672662 | Powdermill |
| *Notiobia nitidipennis* | f | 672661 | Powdermill |
| *Notiophilus aeneus* | m | 819608 | Powdermill |
| *Olisthopus parmatus* | f | 819630 | Powdermill |
| *Olisthopus parmatus* | f | 819633 | Powdermill |
| *Olisthopus parmatus* | f | OSUC 672763 | Powdermill |
| *Olisthopus parmatus* | m | OSUC 672759 | Powdermill |
| *Olisthopus parmatus* | m | OSUC 672760 | Powdermill |
| *Patrobus longicornis* | f | 819693 | Powdermill |
| *Platynus angustatus* | m | 819612 | Powdermill |
| *Platynus angustatus* | m | 819615 | Powdermill |
| *Platynus angustatus* | m | 819618 | Powdermill |
| *Platynus angustatus* | f | 819621 | Powdermill |
| *Platynus angustatus* | f | 819624 | Powdermill |
| *Platynus angustatus* | f | 819627 | Powdermill |
| *Platynus decentis* | m | 819687 | Powdermill |
| *Platynus hypolithos* | m | 819690 | Powdermill |
| *Platynus tenuicollis* | m | 819610 | Powdermill |
| *Platynus tenuicollis* | m | 819607 | Powdermill |
| *Platynus tenuicollis* | f | 819603 | Powdermill |
| *Platynus tenuicollis* | f | 819606 | Powdermill |
| *Platynus tenuicollis* | f | 819609 | Powdermill |
| *Pseudamara arenaria* | f | 819605 | Powdermill |
| *Pseudamara arenaria* | f | OSUC 671695 | Powdermill |
| *Pterostichus adoxus* | m | 819749 | Powdermill |
| *Pterostichus adoxus* | m | 819759 | Powdermill |
| *Pterostichus adoxus* | m | 819769 | Powdermill |
| *Pterostichus adoxus* | f | 819750 | Powdermill |
| *Pterostichus adoxus* | f | 819760 | Powdermill |
| *Pterostichus adoxus* | f | 819770 | Powdermill |
| *Pterostichus atratus* | m | 819735 | Powdermill |
| *Pterostichus atratus* | f | 819736 | Powdermill |
| *Pterostichus coracinus* | m | 819751 | Powdermill |
| *Pterostichus coracinus* | m | 819761 | Powdermill |
| *Pterostichus coracinus* | m | 819771 | Powdermill |
| *Pterostichus coracinus* | f | 819752 | Powdermill |
| *Pterostichus coracinus* | f | 819762 | Powdermill |
| *Pterostichus coracinus* | f | 819772 | Powdermill |
| *Pterostichus corvinus* | m | OSUC 671808 | Powdermill |
| *Pterostichus corvinus* | f | OSUC 671807 | Powdermill |
| *Pterostichus corvinus* | f | OSUC 671809 | Powdermill |
| *Pterostichus corvinus* | f | OSUC 671810 | Powdermill |
| *Pterostichus diligendus* | m | 819781 | Powdermill |
| *Pterostichus hamiltoni* | m | 819779 | Powdermill |
| *Pterostichus lachrymosus* | m | 819682 | Powdermill |
| *Pterostichus lachrymosus* | m | 819679 | Powdermill |
| *Pterostichus lachrymosus* | m | 819676 | Powdermill |
| *Pterostichus lachrymosus* | f | 819673 | Powdermill |
| *Pterostichus lachrymosus* | f | 819670 | Powdermill |
| *Pterostichus lachrymosus* | f | 819667 | Powdermill |
| *Pterostichus melanarius* | m | OSUC 522968 | Erie Co. OH |
| *Pterostichus melanarius* | m | OSUC 522969 | Erie Co. OH |
| *Pterostichus melanarius* | m | OSUC 668451 | Powdermill |
| *Pterostichus melanarius* | f | OSUC 740950 | Cuyahoga Co. OH |
| *Pterostichus moestus* | m | 819737 | Powdermill |
| *Pterostichus moestus* | m | 819741 | Powdermill |
| *Pterostichus moestus* | m | 819745 | Powdermill |
| *Pterostichus moestus* | f | 819738 | Powdermill |
| *Pterostichus moestus* | f | 819742 | Powdermill |
| *Pterostichus moestus* | f | 819746 | Powdermill |
| *Pterostichus mutus* | m | OSUC 522813 | Erie Co. OH |
| *Pterostichus mutus* | m | OSUC 671803 | Powdermill |
| *Pterostichus mutus* | m | OSUC 671804 | Powdermill |
| *Pterostichus mutus* | f | OSUC 522811 | Erie Co. OH |
| *Pterostichus mutus* | f | OSUC 671805 | Powdermill |
| *Pterostichus mutus* | f | OSUC 671801 | Powdermill |
| *Pterostichus rostratus* | m | 819729 | Powdermill |
| *Pterostichus rostratus* | m | 819739 | Powdermill |
| *Pterostichus rostratus* | m | 819743 | Powdermill |
| *Pterostichus rostratus* | f | 819730 | Powdermill |
| *Pterostichus rostratus* | f | 819740 | Powdermill |
| *Pterostichus rostratus* | f | 819744 | Powdermill |
| *Pterostichus sayanus* | m | 819731 | Powdermill |
| *Pterostichus sayanus* | f | 819732 | Powdermill |
| *Pterostichus stygicus* | m | 819755 | Powdermill |
| *Pterostichus stygicus* | m | 819765 | Powdermill |
| *Pterostichus stygicus* | m | 819775 | Powdermill |
| *Pterostichus stygicus* | f | 819756 | Powdermill |
| *Pterostichus stygicus* | f | 819766 | Powdermill |
| *Pterostichus stygicus* | f | 819776 | Powdermill |
| *Pterostichus tristis* | m | 819747 | Powdermill |
| *Pterostichus tristis* | m | 819757 | Powdermill |
| *Pterostichus tristis* | m | 819767 | Powdermill |
| *Pterostichus tristis* | f | 819748 | Powdermill |
| *Pterostichus tristis* | f | 819758 | Powdermill |
| *Pterostichus tristis* | f | 819768 | Powdermill |
| *Scaphinotus andrewsii* | m | 819660 | Powdermill |
| *Scaphinotus andrewsii* | f | 819663 | Powdermill |
| *Scaphinotus imperfectus* | m | 819672 | Powdermill |
| *Scaphinotus imperfectus* | m | 819675 | Powdermill |
| *Scaphinotus imperfectus* | m | 819678 | Powdermill |
| *Scaphinotus imperfectus* | f | 819681 | Powdermill |
| *Scaphinotus imperfectus* | f | 819684 | Powdermill |
| *Scaphinotus ridingsii* | m | 819666 | Powdermill |
| *Scaphinotus ridingsii* | m | 819669 | Powdermill |
| *Scaphinotus viduus* | m | 819654 | Powdermill |
| *Scaphinotus viduus* | m | 819657 | Powdermill |
| *Sphaeroderus canadensis* | m | 819702 | Powdermill |
| *Sphaeroderus canadensis* | m | 819711 | Powdermill |
| *Sphaeroderus canadensis* | f | 819703 | Powdermill |
| *Sphaeroderus canadensis* | f | 819712 | Powdermill |
| *Sphaeroderus canadensis* | f | 819721 | Powdermill |
| *Sphaeroderus stenostomus* | m | 819700 | Powdermill |
| *Sphaeroderus stenostomus* | m | 819697 | Powdermill |
| *Sphaeroderus stenostomus* | m | 819694 | Powdermill |
| *Sphaeroderus stenostomus* | f | 819691 | Powdermill |
| *Sphaeroderus stenostomus* | f | 819688 | Powdermill |
| *Sphaeroderus stenostomus* | f | 819685 | Powdermill |
| *Synuchus impunctatus* | f | 819696 | Powdermill |
| *Trichotichnus autumnalis* | f | 819664 | Powdermill |
| *Trichotichnus autumnalis* | f | 819661 | Powdermill |

Table S\_\_\_\_. Comparison of the principal component analyses run with and without *Notiophilus aeneus*. Traits were listed under “Top loading values” if their loading value was ≥ 0.4.

|  |  |  |
| --- | --- | --- |
|  | **PCA with Notiophilus aeneus** | **PCA excluding Notiophilus aeneus (used in further analyses)** |
| Variance explained by PC1 | 28.9 % | 31.0 % |
| Variance explained by PC2 | 20.6 % | 25.1 % |
| Variance explained by PC3 | 16.4 % | 15.8 % |
| Top loading values for PC1 | pronotum\_width\_standard (-0.55), rear\_trochanter\_length\_standard (-0.46),  rear\_leg\_length\_standard (+0.51) | pronotum\_width\_standard (-0.53), rear\_trochanter\_length\_standard (-0.49),  rear\_leg\_length\_standard (+0.47) |
| Top loading values for PC2 | body\_length (-0.69),  eye\_length\_standard (+0.53) | body\_length (+0.61),  eye\_length\_standard (-0.55),  antenna\_rear\_leg\_ratio (-0.43) |
| Top loading values for PC3 | eye\_protrusion\_ratio (+0.61),  antenna\_rear\_leg\_ratio (-0.55) | eye\_protrusion\_ratio (+0.58),  abdomen\_width\_standard (+0.70) |

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